Genetic polymorphism in dopamine receptor D4 is associated with early body condition in a large population of greater flamingos, *Phoenicopterus roseus*

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Abstract

Body condition is an important determinant of fitness in many natural populations. However, as for many fitness traits, the underlying genes that regulate body condition remain elusive. The dopamine receptor D4 gene (DRD4) is a promising candidate as dopamine is known to play an important role in the regulation of food intake and the metabolism of both glucose and lipids in vertebrates. In this study, we take advantage of a large data set of greater flamingos, Phoenicopterus roseus, to test whether DRD4 polymorphism predicts early body condition (EBC) while controlling for whole-genome effects of inbreeding and outbreeding using microsatellite multilocus heterozygosity (MLH). We typed 670 of these individuals for exon 3 of the homologue of the human DRD4 gene and 10 microsatellite markers. When controlling for the effects of yearly environmental variations and differences between sexes, we found strong evidence of an association between exon 3 DRD4 polymorphisms and EBC, with 2.2-2.3% of the variation being explained by DRD4 polymorphism, whereas there was only weak evidence that MLH predicts EBC. Because EBC is most likely a polygenic trait, this is a considerable amount of variation explained by a single gene. This is to our knowledge, the first study to show an association between exon 3 DRD4 polymorphism and body condition in non-human animals. We anticipate that the DRD4 gene as well as other genes coding for neurotransmitters and their receptors may play an important role in explaining variation in traits that affect fitness.

Keywords: body condition, candidate gene, dopamine receptor, dopamine receptor D4, greater flamingos, multilocus heterozygosity, single-nucleotide polymorphism

Received 20 February 2012; revision received 13 April 2012; accepted 2 May 2012

Introduction

The relative amount of mass in the body not explained by size, that is body condition, is an important determinant of fitness in natural populations (Peig & Green 2009, 2010). In particular, early body condition (EBC) can affect subsequent survival and recruitment into the breeding population in birds (Magrath 1991; Verboven & Visser 1998; Naef-Daenzer *et al.* 2001; Garant *et al.* 2004; Naef-Daenzer & Gruebler 2008), and has been

Correspondence: Mark A. F. Gillingham; E-mail: mark.gillingham@u-bourgogne.fr shown to affect dispersal (Edelman 2011) and the development of foraging skills in mammals (Thornton 2008).

As for almost all quantitative phenotypic traits, EBC variability can be explained by both environmental and genetic factors. However, the identification of genes that underlie EBC, as for most phenotypic variations, remains by and large elusive (Phillips 2005; Mitchell-Olds *et al.* 2007; Mackay *et al.* 2009). This is despite the fact that a major goal for evolutionary biologists is to identify genes that underlie the phenotypic variation in natural populations and to understand the consequences of polymorphism of these loci on fitness (Phillips 2005; Mitchell-Olds *et al.* 2007; Mackay *et al.* 2007; Mackay *et al.* 2009).

A difficult challenge for molecular biologists to overcome is that the link between phenotype and genetic variation is often complex (Phillips 2005; Mackay et al. 2009). Recently, there has been some success in explaining body condition variation using genetic markers by exploring the correlation between body condition or body fat and genetic markers in controlled crosses (QTL analysis) (e.g. Abasht et al. 2006; Moghadam et al. 2007; Rao et al. 2007; Küttner et al. 2011). However, this method is laborious and difficult to achieve in nonmodel organisms that cannot be kept and crossed in laboratory conditions. In addition, QTL approaches have been more successful at identifying genomic regions rather than specific loci which explain phenotypic variation (Phillips 2005; Mackay et al. 2009). Direct genotyping of homologous candidate genes which are known to predict traits in model organisms such as the chicken, mouse and humans that are equivalent to fitness traits in natural populations may provide new insights into our understanding of the relationship between phenotype and genetic variation. Recent studies in molecular ecology have started taking that direction (for a review see Fitzpatrick et al. 2005).

A promising candidate gene is the dopamine receptor D4 (DRD4) gene that codes for one of five types of dopamine receptors in vertebrates (Callier et al. 2003). Neuronal control via various neurotransmitters is known to mediate energy intake and expenditure (for reviews see Meister 2007; Gao & Horvath 2008). Dopamine is one of these important neurotransmitters that regulates many different functions in the central nervous system which in turn affects many traits such as metabolism, food intake, the control of body temperature, and the expression of novelty seeking behaviour (Callier et al. 2003). Dopamine seems to play an important role in regulating food intake by modulating food reward via the mesolimbic circuitry of the vertebrate brain (for reviews see Meister 2007; Gao & Horvath 2008; Wang et al. 2009). Furthermore, glucose and lipid metabolism, two important determinants of weight gain, are strongly affected by dopaminergic neurotransmission. Indeed, in seasonally obese animals, changes in dopaminergic neural activities are known to play a crucial role in the adjustment of body condition (for a review see Pijl 2003).

In vitro studies in humans suggest that a DRD4 allele (the exon 3 DRD4 7-*repeat* allele) has decreased affinity for dopamine and transmits weaker intracellular signals in comparison with other exon 3 alleles (Asghari *et al.* 1995). In addition, in humans, weight gain has been associated with low brain dopamine activity and different exon 3 genotypes of the DRD4 gene are associated with differences in body mass index (BMI; the equivalent of body condition in human studies) and food intake (Levitan *et al.* 2004a,b, 2006; Sobik *et al.* 2005; Guo et al. 2006, 2007; Eisenberg et al. 2008). To what extent DRD4 polymorphism is associated with differences in food intake and expenditure, and therefore differences in body condition, in other vertebrate species is still undocumented. Reflecting the many different functions dopamine seems to regulate, several recent studies have also shown an association between DRD4 polymorphism and variation in novelty seeking and exploratory behaviour, in both humans (for reviews see Savitz & Ramesar 2004; Ebstein 2006) and non-human vertebrates (Momozawa et al. 2005; Bailey et al. 2007; Fidler et al. 2007; Hejjas et al. 2007; James et al. 2007; Flisikowski et al. 2009; Korsten et al. 2010). Such pleiotropism of the DRD4 gene further validates it as a promising candidate gene to explain variance in body condition because individual variation in behaviour is expected to covary with other life history, physiological and metabolic traits (for reviews see Biro & Stamps 2008, 2010; Houston 2010; Réale et al. 2010).

In this study, we take advantage of a large data set of greater flamingos, Phoenicopterus roseus, to test whether DRD4 polymorphism predicts EBC. As in many birds, EBC has been found to be an important predictor of postfledging dispersal in flamingos (Barbraud et al. 2003). Because postfledging dispersal is an important ecological process with numerous fitness implications (Hamilton & May 1977; Comins et al. 1980; Greenwood & Harvey 1982; Clobert et al. 2001), EBC may be an important predictor of fitness in flamingos. Indeed, Sanz-Anguilar et al. (in press) have found that survival differed according to dispersal strategies in greater flamingos. Long-distance dispersers survival was lower than residents or intermediate-distance dispersers for one- and two-year old birds, while the opposite trend was found for older birds (Sanz-Aguilar et al. in press).

The deleterious effects of inbreeding and outbreeding on the whole genome may also have an effect on phenotype variation (David et al. 1995; Coltman & Slate 2003; Chapman et al. 2009; Szulkin et al. 2010), including body condition (e.g.: Lieutenant-Gosselin & Bernatchez 2006). If inbreeding is recurrent in the population then theory predicts a correlation in heterozygosity and/or homozygosity across loci, known as identity disequilibrium (ID) (Szulkin et al. 2010). In addition, if there is admixture between populations (outbreeding), alleles at two loci may be preferentially associated in gametes [linkage disequilibrium (LD)] and the random association of gametes yields an excess of double-heterozygous genotypes (therefore also resulting in ID) (Szulkin et al. 2009). Thus, if inbreeding and outbreeding are strong in the population, microsatellite multilocus heterozygosity (MLH) may reflect genome-wide heterozygosity as a result of ID (Coltman & Slate 2003; Chapman et al. 2009; Szulkin et al. 2010). Associations between MLH and fitness traits are known as heterozygosity-fitness correlation (HFC) (Coltman & Slate 2003; Chapman *et al.* 2009; Szulkin *et al.* 2010). To control for any potential artefactual effects of DRD4 polymorphism on EBC that may be due to inbreeding and/or outbreeding, we also explore the statistical association between MLH and EBC and between MLH and DRD4 heterozygosity.

Long-term monitoring of the Camargue population of greater flamingos (Johnson & Cézilly 2007) has revealed that environmental conditions, such as water levels around the breeding colony, significantly affect chick body condition (Cézilly *et al.* 1995; Béchet & Johnson 2008). Another important factor that may influence body condition is differential allocation of nutritional resources towards growth and storage between males and females. Therefore, we statistically controlled for the effects of both sex and annual environmental conditions (over four consecutive years) when investigating whether DRD4 polymorphism influences EBC.

Methods

Study area and species

Greater flamingos have bred every year at the Fangassier's lagoon (43°25'N, 4°37'E; Salin de Giraud, Camargue, southern France) since 1969 (except in 2007; Béchet et al. 2012). Between 1995 and 1998, blood samples were collected from chicks during ringing operations in July/August and preserved in a blood buffer (Seutin et al. 1991). Chicks were caught on average 106 days after the start of laying just before the oldest chicks fledged, by herding the crèche into a corral. Thus, a random sample of chicks, which included early- and late-hatching birds, were captured (approximate range 35-77 days) (Johnson & Cézilly 2007). During the ringing operation, chicks were marked individually with PVC plastic rings engraved with a four-digit code and measured (tarsus length to the nearest 1 mm and body weight to the nearest 50 g using a 5-kg Pesola spring balance; see Johnson & Cézilly (2007) for details). Descriptive statistics of mean chick weight and tarsus length according to sex and cohort year are provided in Table 1. As greater flamingos only lay one egg per season and switch mates systematically between consecutive breeding seasons, there is no sibship relationships in this study (Cézilly & Johnson 1995; Johnson & Cézilly 2007).

DNA extraction and sexing

DNA was extracted using a standard phenol-chloroform method (Hillis *et al.* 1996). The quality and con**Table 1** Mean and standard deviation of chick weight (g) and chick tarsus length (mm) according to cohort and sex

Year	Sex	Sample size	Mean weight (SD)	Mean tarsus length (SD)
1995	Female	68	1877 (348)	198 (20)
	Male	60	2152 (432)	216 (21)
1996	Female	40	2673 (343)	228 (13)
	Male	38	2869 (411)	243 (20)
1997	Female	127	2311 (331)	210 (15)
	Male	132	2585 (383)	221 (17)
1998	Female	120	2044 (411)	199 (21)
	Male	85	2303 (434)	214 (22)
Total		670	2311 (472)	213 (22)

centration of DNA extracted was estimated by UV spectrophotometry (Spectramax plus 384, Molecular devices) to ensure that a final dilution of approximately 50 ng/ μ L was achieved. Molecular sexing was based on CHD gene sequences polymorphism and was carried out as described by either Bertault *et al.* (1999) or Balkız *et al.* (2007).

DRD4 genotyping

Six hundred and seventy individuals belonging to four cohorts: 1995 (*n* = 128), 1996 (*n* = 78), 1997 (*n* = 259) and 1998 (n = 205) were sequenced for DRD4 exon 3. DRD4 exon 3 sequences from Gallus gallus DRD4 (NM001142849 and FJ217173), Parus major (DQ006802) and Taeniopygia guttata (GQ359780) were aligned manually using MEGA version 5 (Tamura et al. 2011) allowing the definition of two degenerated primers: F1-E3-DR4D (5'-CCRCTSAACTACAACCGGCG-3') and R1-E3-DR4D (5'-YTCCCGGCCGTTGATCTTGG-3'). These primers amplify 486bp of DRD4 exon 3. Polymerase chain reactions (PCRs) were performed with approximately 40 ng of extracted DNA in 25 µL reactions containing 50 µM of each dNTP (Euromedex), 200 nm of each primer, 0.15 units of Manual HotMaster[™] Taq DNA polymerase (5') and 2.5 µL of the supplied 10× buffer. PCR programme comprised an initial denaturation step of 3 min at 95 °C, 35 cycles at 94 °C for 45 s, 60 °C for 60 s and 65 °C for 60 s, followed by a final extension step at 65 °C for 5 min. Prior to direct sequencing PCR products were purified by using exonuclease 1 and shrimp alkaline phosphatase (Fermentas Life Sciences). Products were then sequenced using the forward primer F1-E3-DR4D at the Macrogen Sequencing Service (Macrogen Inc., South Korea). Sequence editing, BLAST searches and multiple sequence alignments carried out using clustalw were executed in MEGA. Six synonymous singlenucleotide polymorphism (SNP) sites were identified.

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A total of 23 DRD4 exon 3 genotypes were identified at varying frequencies (Fig. 1a).

Thirteen heterozygote individuals (Fig. 1a) encompassing all putative alleles were cloned to enable the characterization of every DRD4 exon 3 allele. Amplicons were cloned using the pGEM-T Easy Vectors (Promega) kit and JM109 competent cells (Promega) by following manufacturers' guidelines. At least seven clones were sequenced from each individual and each allele was sequenced in at least two individuals. Eight alleles, named a to h (GenBank Accession entries for alleles are JQ425594, JQ425595, JQ425596, JQ425597, JQ425598, JQ425599, JQ425600 and JQ425601, respectively), were identified from cloned sequences. Genotypes of all remaining directly sequenced individuals were inferred based on a combination of these eight alleles, allowing the definition of the 23 genotypes. Although we cannot be certain that all alleles were identified, the fact that all of the eight alleles identified (each sequenced in at least two individuals) allowed the definition of the 23 observed genotypes would suggest that they were. Furthermore, when analysing the association of DRD4 polymorphism and EBC, we used only the five most common genotypes (see below), which were either cloned (ac and bc), homozygotes (aa and bb) or a heterozygote genotype of the two most common alleles in the



Fig. 1 Proportion of greater flamingos in the sampled population with DRD4 exon 3 genotype (a) and DRD4 exon 3 allele (b). Lower case font represents DRD4 exon 3 alleles and number of individuals with DRD4 genotype and alleles are indicated within brackets. An individual was chosen for cloning for genotypes marked with an *. population (ab). Deviations from Hardy–Weinberg equilibrium were tested with Genepop 4.0.10. (Rousset 2008).

Microsatellite genotyping

Genotyping of microsatellite markers was performed at 10 microsatellite loci (PrD3, PrD4, PrD5, PrD9, PrA110, PrA113, PrC109, PrD108, PrD121 and PrD126) developed for the Greater flamingo (An et al. 2010) (PCRs conditions can be found at http://tomato.bio.trinity.edu/manuscripts/10-2/mer-09-0396.pdf; GenBank Accession entries for microsatellites are GF101824, GF101825, GF101826, GF101828, GF101835, GF101837, GF101842, GF101847, GF101892 and GF101849 respectively). Electrophoresis was performed on a 96 capillary sequencer ABI3730XL (GENTYANE, INRA, France). Alleles were scored using GeneMapper 4.0 (Applied Biosystems). For each cohort, the program MICRO-CHECKER (Van Oosterhout et al. 2004) was used to test for genotyping errors. Identity disequilibrium, predicted to occur whether inbreeding is common in a population, was estimated from parameter g_{2} , using the RMES software available at http://ftp.cefe.cnrs.fr (David et al. 2007). A significant departure of g_2 from 0 is a powerful way of detecting ID in the population (David et al. 2007; Szulkin et al. 2010).

Multilocus heterozygosity

Multilocus heterozygosity was measured as the sum of heterozygote loci divided by the number of loci (Chapman *et al.* 2009; Szulkin *et al.* 2010). We also calculated three other measures of MLH, namely internal relatedness (IR, Amos *et al.* 2001), standardized heterozygosity (SH, Coltman *et al.* 2001), standardized heterozygosity by loci (HL, Aparicio *et al.* 2006) using the R function GEN-HET (Coulon 2010). All MLH measures were highly correlated and results were highly similar to analyses using MLH (Appendix S1, Supporting information). We only present here the results using MLH following recommendations from Chapman *et al.* (2009).

Calculating body condition

When estimating body condition, chick tarsus length was used as a body size indicator as in previous studies (Cézilly *et al.* 1995; Barbraud *et al.* 2003; Béchet & Johnson 2008), which significantly correlates with chicks' mass [standardized major axis (SMA) regression of the log-transformed mass–length relationship; $R^2 = 0.72$; P < 0.001, slope = 2.01]. We used the scaled mass index to calculate EBC (Peig & Green 2009, 2010), in order to control for the effects of body size on both the independent and dependent variables. In addition, the scale

mass index retains the original units of measurement. However in our study, using more traditional methods of measuring body condition, that is, the residuals from an ordinary least squares (OLS) or SMA regressions of the log-transformed mass-length relationship, were highly collinear with the scale mass index and results were highly similar to analyses using the scale mass index (Appendix S2, Supporting information). Student residuals from an OLS regression of log-transformed mass and log-transformed tarsus length were used to test for significant outliers with the R function CAR in the software package R-2.14.2 (http://www.R-project.org). Three significant outliers (Bonferroni adjusted P < 0.05) were removed from the data set (from a total of 670 individuals) and were considered to be the result of measurement errors at ringing.

When calculating the scaled mass index, we used the slope of the SMA of log-transformed mass against log-transformed tarsus length for all cohorts and both sexes confounded as the scaling exponent using the R function SMATR. By confounding all cohorts and sex, we can test whether EBC varied significantly between cohorts (1995, 1996, 1997 and 1998) and sexes. This was legitimate because of a lack of difference in slopes (P > 0.05) when fitting a SMA regression of log-transformed mass against log-transformed tarsus within each cohort and sex. Therefore, despite morphological differences between sexes and cohorts, we assumed the same morphogenetic pattern for males and females between each cohort.

Statistical analysis

All statistical analyses were carried out using the software package R-2.14.2 (http://www.R-project.org). To investigate the effect of DRD4 polymorphisms on EBC, genotypes that were found in fewer than 20 individuals were excluded from the data set, thus avoiding false interpretation because of small sample sizes (type I and type II errors). As a result, only five genotypes (aa, bb, ab, ac and bc) and three alleles (a, b and c) were considered when analysing the association of DRD4 genotypes with EBC. We chose to investigate whether DRD4 genotypes rather than individual SNPs predict EBC because of a lack of independence between SNPs within DRD4 exon 3 (Appendix S4, Supporting information). However, using SNPs instead of DRD4 genotypes gave virtually identical results (Appendix S4, Supporting information). Therefore, 581 of 670 individuals genotyped for DRD4 exon 3 were included in this analysis. Model selection was achieved through information-theoretic (I-T) model selection and multimodel inference approach (Burnham & Anderson 2002), because recent literature recommends using this statistical approach

for observational studies (see the following reviews for I-T model selection in behavioural ecology: Johnson & Omland 2004; Garamszegi et al. 2009; Hegyi & Garamszegi 2011; Richards et al. 2011; Symonds & Moussalli 2011). Burnham & Anderson (2002) provide a complete description of model selection methods. All possible candidate models were constructed using the predictor variables cohort (1995, 1996, 1997 or 1998), sex (female or male), DRD4 genotype (aa, bb, ab, ac or bc) and the two-way interaction between sex and cohort. As both inbreeding and outbreeding may generate HFC, a complex relationship between MLH and body condition can be expected if effects are pulled by extreme values of MLH (i.e. among the very inbred and/or very outbred individuals). Therefore, we initially tested for a nonlinear association between MLH and body condition using general additive modelling (GAM) fitted with a Gaussian distribution with the R package MGCV (Wood & Augustin 2002), which automatically tests for nonlinear response shapes by fitting smoothed functions for the predictor variables (Guisan et al. 2002; Zuur et al. 2009). MLH was fitted with a smoothed function with an upper limit of five degrees of freedom. As all GAM models did not report significant nonlinear relationships and gave equivalent Akaike's Information Criterion (AIC) to linear models, we continued model selection using linear regression models and concluded that there was no evidence of a nonlinear association between MLH and body condition. AIC weights (ω) were used to assess the relative strength of support for models (Burnham & Anderson 2002; Johnson & Omland 2004). Only the 10 top ranked models based on AIC are presented. Parameter estimates were calculated using model averaging whereby regression coefficients are based on the full suite of models described previously. This procedure is more robust when several models have similar support (Burnham & Anderson 2002; Johnson & Omland 2004). The relative importance of each predictor variable is estimated by summing the AIC weight in which that variable appears across supported models ($\Sigma AIC\omega$) (Burnham & Anderson 2002). A summed Akaike weight value tends towards 1 if a particular predictor appears in all of the top models. Conversely, a summed Akaike weight value tends towards 0 if a particular predictor appears only in models with low support (Burnham & Anderson 2002; Symonds & Moussalli 2011).

Heterozygosity-fitness correlation may be generated independently from inbreeding and outbreeding if there is overdominant linkage between one or more microsatellite loci to one or more functional molecular markers [referred to as a local effect (Szulkin *et al.* 2010)]. Although, as argued by Szulkin *et al.* (2010), this is highly unlikely particularly if HFC is weak (as is the

association between MLH and EBC, see results), we nonetheless tested for a significant local effect of MLH using the methods described by Szulkin et al. (2010). Briefly, an *F*-ratio test was used to test whether a model containing each single loci heterozygosity (SLH; expressed as 0 or 1, missing values replaced by the mean at that locus) explained more of the variance than a model containing MLH (calculated for this test as the sum of SLH, missing values replaced by the mean at that locus). The best-ranked model with MLH (model 2 in Table 2a) was used for the local effects test. Thus, a model containing the predictor variables cohort, sex, DRD4 genotypes and MLH was compared to a model with predictor variables cohort, sex, DRD4 genotypes and the 10 SLH to test for significant local effects. We used the top ranked model (model 1 in Table 1a) to test whether DRD4 heterozygosity (DRD4 homozygote or heterozygote) rather than DRD4 genotype predicted EBC. We also evaluated whether MLH predicted DRD4 heterozygosity using a GLM with a binomial distribution, DRD4 heterozygosity as the response variable and MLH as the explanatory variable.

To check whether a significant association in DRD4 polymorphism was not driven by rare genotypes, the analysis was repeated after excluding all genotypes with fewer than 99 individuals. As a result, only three genotypes (aa, bb and ab) and two alleles (a and b) were considered, which represents 79% of the sampled greater flamingo population. Consequently, 526 of 670 individuals genotyped for DRD4 exon 3 were included in this second analysis.

Results

DRD4 exon 3 genotyping

Amplification of exon 3 DRD4 was confirmed through BLASTP search of the GenBank (nr database) using the predicted 161 residue greater flamingo DRD4 protein which returned *E*-values of $4 \times e^{-99}$, $2 \times e^{-98}$, $7 \times e^{-98}$, $1 \times e^{-96}$ and $2 \times e^{-94}$ for alignments with the chicken, *Gallus gallus* (NP001136321), the wild turkey, *Meleagris gallopavo* (XP003206209), the blackcap, *Sylvia atricapilla* (AEC22814), the zebra finch, *Taeniopygia guttata* (ACT99861) and the great tit, *Parus major* (AAY56686) DRD4 protein sequences, respectively. The DRD4 exon 3 protein showed 87% identity with the chicken DRD4 protein, 63% with the mouse DRD4 protein and 61% with the human DRD4 protein.

A total of six synonymous SNPs were identified, defining eight alleles (named a–h), these alleles being combined in 23 observed genotypes (Fig. 1). There was no evidence that DRD4 genotypes significantly deviated from Hardy– Weinberg equilibrium (n = 670; *P*-value = 0.1673;

Table 2 Top 10 models of early body condition of greater flamingos with the five most common DRD4 exon 3 genotypes included in the data set (a; n = 581) and the three most common DRD4 exon 3 genotypes included in the data set (b; n = 526), showing number of parameters (k), log-likelihood (LL), AIC of the models, change in AIC compared with the best-ranked model (Δ AIC), and Akaike model weights (ω). The base model included sex (male or female), cohort, microsatellite heterozygosity, DRD4 genotype and the interaction between sex and cohort

Model rank	Model	k	LL	AIC	Δ AIC	ω
(a) Models wit	th only the five most common DRD4 genotypes ($n = 581$)					
1	Cohort + DRD4 genotype + Sex	10	-3955.29	7930.6	0	0.287
2	Cohort + DRD4 genotype + Sex + MLH	11	-3954.50	7931.0	0.43	0.231
3	Cohort + DRD4 genotype + Sex + Cohort \times Sex	13	-3952.77	7931.5	0.96	0.177
4	Cohort + DRD4 genotype + Sex + Cohort × Sex + MLH	14	-3952.02	7932.0	1.47	0.138
5	Cohort + DRD4 genotype	9	-3957.81	7933.6	3.05	0.062
6	Cohort + DRD4 genotype + MLH	10	-3956.97	7933.9	3.36	0.054
7	Cohort + MLH + Sex	7	-3961.05	7936.1	5.52	0.018
8	Cohort + Sex	6	-3962.09	7936.2	5.6	0.017
9	$Cohort + Sex + Cohort \times Sex + MLH$	10	-3959.45	7938.9	8.32	0.004
10	$Cohort + Sex + Cohort \times Sex$	9	-3960.47	7938.9	8.37	0.004
(b) Models wit	th only the three most common DRD4 genotypes ($n = 526$)					
1	Cohort + DRD4 genotype + Sex	8	-3582.46	7180.9	0	0.257
2	Cohort + DRD4 genotype	7	-3583.91	7181.8	0.88	0.165
3	Cohort + DRD4 genotype + Sex + MLH	9	-3582.10	7182.2	1.27	0.137
4	Cohort + DRD4 genotype + Sex + Cohort \times Sex	11	-3580.37	7182.7	1.8	0.104
5	Cohort + DRD4 genotype + MLH	8	-3583.55	7183.1	2.17	0.087
6	Cohort + Sex	6	-3585.84	7183.7	2.76	0.065
7	Cohort + DRD4 genotype + Sex + Cohort × Sex + MLH	12	-3579.95	7183.9	2.97	0.058
8	Cohort	5	-3587.35	7184.7	3.78	0.039
9	Cohort + Sex + MLH	7	-3585.52	7185.0	4.11	0.033
10	$Cohort + Sex + Cohort \times Sex$	9	-3583.90	7185.8	4.87	0.023

SE = 0.0196). DRD4 genotypes were found at an uneven frequency with for example the heterozygote genotype ab present in almost 40% of birds, and the homozygote genotypes aa and bb in 24% and 15% of birds, respectively (Fig. 1a). Alleles a and b were present in almost 75% and 63% of birds, respectively, with the next most common allele c present in only 9% of birds (Fig. 1b). None of the alleles differed by more than four base pairs, with a mean of 2.1 and a standard deviation of 0.91 (Appendix S3, Supporting information). Furthermore, the three most common alleles (a, b and c) that defined the five most common genotypes (aa, bb, ab, ac and bc) only differed by two base pairs (from three SNPs; Appendix S3, Supporting information).

Association between EBC and DRD4 polymorphism

Four models were retained as equivalent (Δ AIC < 2), all of which retained an effect of DRD4 polymorphism, cohort and sex (Table 2 a). Model averaging revealed very strong support of an association between DRD4 genotypes and EBC (Σ AIC ω = 1; Table 3a) and DRD4 genotypes explained between 2.2 and 2.3% of EBC variance according to the 4 equivalent models. Chicks carrying genotypes aa, bc and ab were of lower body condition than those carrying genotypes ac and bb, with individuals of genotype ac in higher body condition than those of genotype bb (Fig. 2). Indeed, EBC of genotypes ac and bb were on average 5.7% and 2.8% higher, respectively, than individuals carrying genotype aa (the genotype with the lowest mean adjusted EBC; Fig. 2). Chicks carrying genotypes aa, bc and ab were of similar condition (Fig. 2). Virtually identical results were found when using individual DRD4 SNPs, whereby there was strong evidence of an association between EBC and the three DRD4 exon 3 SNPs (although there was a lack of independence between different SNPs; Appendix S4, Supporting information). These combined results are suggestive of a complex interaction between alleles rather than a simple allele dominance interaction. Model averaging also revealed very strong support of variation in EBC between cohorts ($\Sigma AIC\omega = 1$; Table 3a; Fig. 3) and strong support of higher EBC in females compared with males $(\Sigma AIC\omega = 0.88; Table 3a; Fig. 3)$. There was only weak support for an interaction between sex and cohort $(\Sigma AIC\omega = 0.33; Table 3a; Fig. 3).$

No significant among-locus correlation between microsatellite loci was detected ($g_2 = -0.00004$, SD = 0.002, P = 0.520). However, a significant association between

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Table 3 Model-averaged parameter estimates of early body condition of greater flamingos with individuals with the five most com-
mon DRD4 exon 3 genotypes included in the data set (a; $n = 581$) and the three most common DRD4 exon 3 genotypes included in
the data set (b; $n = 526$). Summed AIC weight of each parameter is also shown ($\Sigma AIC\omega$). See Table 3 for descriptions of models

Parameter	Model-averaged estimate	Adjusted SE	95% C.I.	ΣΑΙCω		
(a) Data set with the five most co	ommon DRD4 genotypes					
Intercept (females, 1995, aa)	2093.31	52.60	1990.22, 2196.40			
Cohorts				1		
1996	191.04	44.08	104.64, 277.44			
1997	274.15	31.74	211.95, 336.35			
1998	193.00	31.57	131.13, 254.87			
Sex				0.88		
Males	-49	31.36	-110.47, 12.47			
Cohorts×Sex:				0.33		
1996 (Males)	-80.04	68.33	-213.97, 53.88			
1997 (Males)	49.67	51.38	-51.04, 150.38			
1998 (Males)	37.70	53.84	-67.83, 143.23			
DRD4 genotypes				1		
ab	-0.38	22.25	-44, 43.23			
ac	131.21	47.67	37.78, 224.64			
bb	63.91	28.28	8.48, 119.35			
bc	17.17	44.81	-70.64, 104.99			
MLH	-82.38	66.64	-213, 48.23	0.45		
(b) Data set with the three most	common DRD4 genotypes					
Intercept (females, 1995, aa)	2072.86	46.58	1981.57, 2164.16			
Cohorts				1		
1996	195.03	43.74	109.30, 280.76			
1997	283.42	29.91	224.80, 342.04			
1998	196.08	30.86	135.59, 256.56			
Sex				0.69		
Males	-36.97	28.47	-92.77, 18.83			
Cohorts×Sex:				0.17		
1996 (Males)	-90.79	74.11	-236.05, 54.47			
1997 (Males)	37.74	53.62	-67.36, 142.83			
1998 (Males)	36.05	56.21	-74.11, 146.21			
DRD4 genotypes						
ab	-1.09	22.30	-44.79, 42.61			
bb	63.99	28.37	8.38, 119.59			
MLH	-60.88	71.57	-201.14, 79.39	0.33		

MLH and EBC may still occur if significant ID is not detected because a slight inbreeding is often more easily detected through its effects on phenotype than through its effect on heterozygosity at a few marker loci (Szulkin et al. 2010). However, evidence that MLH predicted EBC was weak ($\Sigma AIC\omega = 0.45$, Table 3a). MLH was not retained in the best-ranked model (Table 2a), and there was weak support for the slope of the term being different from zero (Table 3a). Furthermore, including MLH in the model did not change the strong association of DRD4 polymorphism with EBC. Testing for significant local effects on body condition was achieved as advised by Szulkin et al. (2010) and revealed no significant evidence of local effects on EBC ($F_{19,552} = 1.081$, P = 0.366). DRD4 homozygotes had a significantly lower MLH than DRD4 heterozygotes ($\chi^2_{1,579} = -4.902$; P = 0.027), but DRD4 heterozygosity did not significantly predict EBC ($F_{1,575} = 0.327$; P = 0.568).

The analysis of the association of DRD4 genotypes with EBC was repeated with only the three most common DRD4 genotypes aa, bb and ab in the analysis, which represents 79% of the sampled greater flamingo population (at least 99 individuals per DRD4 genotype). Again DRD4 polymorphism was retained by all of the equivalent models (Table 2b), with 1.3% of the variance explained by DRD4 genotypes according to the four equivalent models ($\Sigma AIC\omega = 0.86$; Table 3b).

Discussion

We found strong evidence that EBC in the Greater flamingo was associated with DRD4 exon 3 polymorphism



Fig. 2 Adjusted mean early body condition (±SEM) of greater flamingos according to the five most common DRD4 genotypes (corrected for cohort and sex effects using model 1 in Table 2). Sample sizes are indicated within brackets.



Fig. 3 Adjusted mean early body condition (\pm SEM) of greater flamingo chicks in relation to cohort and sex (corrected for DRD4 genotypes effect using model 3 in Table 2). Sample sizes are indicated within brackets.

but not with MLH. As expected, we also observed strong evidence of variation in EBC between cohorts that reflects the effects of between-year stochastic environmental conditions. Annual variation (as encompassed by the cohort effect) in EBC explained between 17.3 and 17.6% of the variance (according to the four equivalent models) in our study. This is consistent with previous studies which seem to show that water levels around the breeding colony and number of breeding pairs have a significant effect on EBC (Cézilly et al. 1995; Béchet & Johnson 2008). Interestingly, we found for the first time in this species a significant difference in EBC between females and males, with females in a significantly higher condition than males. In our study, between 0.8 and 0.9% (according to the four equivalent models) of the variance in EBC was explained by differences between males and females. Although it is known that there is morphological sexual dimorphism in adult flamingos with males tending to be larger than females (Johnson & Cézilly 2007), no previous study had reported differential body condition between the sexes.

When excluding DRD4 exon 3 genotypes with fewer than 20 individuals, 2.2-2.3% of the variation in EBC was explained by DRD4 polymorphisms (although this increased to 4.9% when all genotypes were included in the analysis). Differences in EBC between the two most extreme DRD4 exon 3 genotypes were in the order of 6%. Molecularly this seems to be a substantial effect especially when considering that, as is the case for behavioural traits, body condition is probably to be regulated by multiple genes (reviewed in Snyder et al. 2004). Indeed, a minimum of five independent association studies (reviewed in Snyder et al. 2004) have provided evidence for at least 15 candidate genes which are probably to regulate body composition in humans. As a point of comparison, in a study in humans investigating the association between changes in body fatness over time and 15 polymorphisms in 10 candidate genes of obesity (although not DRD4 polymorphism in this study), Bouchard et al. (2007) found that only 3-5 genes were retained in the model with the strongest candidate gene only explaining 3.3% of the variance. Furthermore, strong evidence of an association between DRD4 polymorphism and EBC remains even when genotypes with fewer than 99 individuals are removed from the analysis. Therefore, although EBC is probably mediated by several genes, our results do provide robust correlative evidence that DRD4 polymorphism, or a gene closely linked to the DRD4 gene, is associated with it. A similar result was found when investigating the association between exploratory behaviour and exon 3 DRD4 polymorphism in a Dutch population of great tits (Fidler et al. 2007; Korsten et al. 2010). A single synonymous SNP in exon 3 of DRD4 was associated with exploratory behaviour explaining 4.5–5.8% of the variation, although this association was not present in three other wild European populations (Fidler et al. 2007; Korsten et al. 2010).

Multilocus heterozygosity from a few microsatellites markers is often poorly correlated with pedigree-based values of inbreeding coefficient F (Szulkin et al. 2010). We cannot exclude the possibility that a lack of association between MLH and EBC may be due to a lack of power in detecting inbreeding depression because of the relatively small number of markers (10) used in this study. Indeed we found that individuals that were DRD4 heterozygotes had significantly lower MLH values than DRD4 homozygotes. This contradictory result may be further suggestive of a lack of power in detecting inbreeding. However, alternatively MLH may have been a poor predictor of whole-genome heterozygosity because the greater flamingo population may be a large panmictic population at equilibrium and at low risk of the effects of inbreeding and outbreeding depression. A recent study on a large (n = 1192), moderately inbred, captive population of zebra finch (Taeniopygia guttata) compared the power of 11 microsatellite markers, 1359 SNPs markers and pedigree-based F in terms of detecting inbreeding depression on 11 phenotypic traits (Forstmeier et al. in press). The authors found that microsatellite and SNP markers produced equally strong HFCs and that both markers produced stronger HFCs than the pedigree-based F. Forstmeier et al. (in press) concluded that a small panel of microsatellites may reflect better an individual's realized inbreeding depression than previously appreciated. Therefore. our results do at least suggest that the inbreeding and outbreeding effects may be relatively unimportant in explaining the associations between the DRD4 genotypes and EBC. Moreover, EBC was associated with specific DRD4 genotypes rather than DRD4 heterozygosity.

Previous studies in humans have found a significant association between DRD4 exon 3 polymorphism and body condition but to our knowledge this is the first study to find such an association in other vertebrates. In human studies, associations between DRD4 polymorphisms and body condition involve nonsynonymous polymorphisms within exon 3, with studies focusing on a 48 bp repeat (Levitan et al. 2004a,b, 2006; Sobik et al. 2005; Guo et al. 2006, 2007; Eisenberg et al. 2008). In our study, all six SNPs were synonymous. However, as argued by Fidler et al. (2007) and Korsten et al. (2010), a significant association between synonymous SNPs at exon 3 of DRD4 and a phenotypic trait may be biologically meaningful for two reasons. First, synonymous polymorphism within exon 3 may be linked to other polymorphisms either within other exon or promoter regions of the DRD4 gene (e.g. exon 1 which codes for the extracellular domain) and/or to other genes that may also regulate body condition. For example, in chickens, it appears that DRD4 is in LD with deformed epidermal auto regulatory factor one (DEAF1), a gene

involved in the regulation of the serotonergic system (Flisikowski et al. 2009). Second, there is now a growing consensus that synonymous nucleotide substitutions may alter protein function and can be targeted by natural selection (for a review see Sauna & Kimchi-Sarfaty 2011). Indeed, it has been shown that there is codon usage bias between synonymous codons (Chamary et al. 2006; Kimchi-Sarfaty 2007; Plotkin & Kudla 2011). It has recently been demonstrated that codon bias because of silent substitutions can affect protein conformation and have functional consequences (Chamary et al. 2006; Kimchi-Sarfaty 2007; Plotkin & Kudla 2011). Moreover, it has been demonstrated that synonymous polymorphisms influence mRNA stability and translation of dopamine receptors (Duan et al. 2003). Therefore, synonymous substitutions in DRD4 exon 3 may cause functional differences in protein, changing affinity to dopamine which in turn results in different EBC between genotypes.

Assuming that DRD4 exon 3 polymorphism does result in a differential dopamine affinity in the greater flamingo, we propose three nonmutually exclusive hypotheses that might explain differences in EBC between DRD4 genotypes. First, food intake and energy homoeostasis may be different between DRD4 genotypes. Indeed, dopamine is thought to regulate body weight in vertebrates by playing an important role in the motivational mechanisms associated with the behavioural responses necessary for food intake (for reviews see Meister 2007; Gao & Horvath 2008; Wang et al. 2009) as well as the metabolism of glucose and lipids (for a review see Pijl 2003). Second, chick begging behaviour may differ according to exon 3 DRD4 genotype. Recent studies indicate that behavioural types, such as bold and aggressive personalities, are positively related to food intake rates (reviewed in Biro & Stamps 2008) and energy expenditure (Careau et al. 2010, 2011). Considering that recent studies suggest a significant association between personalities and DRD4 polymorphism (Momozawa et al. 2005; Bailey et al. 2007; Fidler et al. 2007; Hejjas et al. 2007; James et al. 2007; Flisikowski et al. 2009; Korsten et al. 2010), certain DRD4 genotypes might be bolder and more aggressive during begging, resulting in higher food intake and, in turn, higher EBC. However, it is important to emphasize that these potential differences in food intake because of begging behaviour do not result from differences in competitive ability between siblings because greater flamingos only lay one egg per season. Third, an association between EBC and DRD4 genotype may result from an indirect effect of differences in parental foraging behaviour between parental DRD4 exon 3 genotypes. In this scenario, parental rather than chick DRD4 genotypes would be a better predictor of EBC.

Somewhat paradoxically the DRD4 genotypes that were associated with higher EBC were not the most common in the population. Considering the pleiotropic nature of the DRD4 gene, it is possible that DRD4 genotypes associated with higher EBC may have other fitness consequences. Moreover, to what extent differences in mean EBC between DRD4 exon 3 genotypes affect fitness remains to be tested. Future studies should therefore investigate whether DRD4 polymorphisms also predicts survival and recruitment into the breeding population. Recent studies in humans indicate that there is an association between migration and exon 3 DRD4 polymorphisms (Chen et al. 1999; Matthews & Butler 2011). Interestingly, Barbraud et al. (2003) found that EBC is an important predictor of natal dispersal. Thus, future studies should also focus on investigating a link between dispersal and DRD4 polymorphism in the greater flamingo. So far, a single study has investigated if DRD4 polymorphism is associated with migration (but not dispersal) in non-human animals (blackcaps, Sylvia atricapilla) and found no effect (Mueller et al. 2011).

Acknowledgements

We thank Maria Teixeira Brandao, Charles Poncet, Lydia Jaffrelo, Nathalie Bernard, and Pierre Desray for their invaluable assistance in the laboratory. Salins Group kindly granted access to the flamingo colony in the Camargue. This study was funded by the TOTAL foundation, the Conseil Régional de Bourgogne, the MAVA foundation, and the Centre National de la Recherche Scientifique (CNRS). Mark Gillingham was supported by postdoctoral grant for the Conseil Régional de Bourgogne. Julia Geraci was supported by a doctoral grant cofunded by La Tour du Valat and the Conseil Régional de Bourgogne. We are deeply grateful to Luc Hoffmann and Alan Johnson for the instigation of the long-term study on the Greater flamingo and to Christophe Germain, Antoine Arnaud, and Michel Gauthier-Clerc from the Tour du Valat for special support to this study. We are also grateful to François-Xavier Dechaume-Moncharmont, Caroline Zanchi and three anonymous reviewers for comments on previous drafts.

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Data accessibility

DRD4 genotypes, microsatellite genotypes, MLH and EBC: DRYAD entry doi:10.5061/dryad.rd45qj52

Supporting information

Additional supporting information may be found in the online version of this article.

Appendix S1. Correlation between multilocus heterozygosity and other measures of microsatellite multilocus heterozygosity measures.

Appendix S2. Correlation between early body condition estimated as the scale mass index against residuals of an SMA and OLS regression of log (mass) and log (tarsus).

Appendix S3. Mutational distance between different DRD4 alleles.

Appendix S4. Early body condition according to DRD4 SNPs.

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